

CO₂ extreme dissolved concentrations influence on growth and CHO cells metabolic characteristics in periodic and continuous processes

© Dmitry V. Tyupa,^{1*} Anton N. Morozov,^{1*} Zakhar V. Zakharov,¹
Sergey V. Kalenov,²⁺ Roman A. Kochelabov,¹ and Ilia M. Emelyanov¹

¹ IBC “Generium”. Vladimirskaya St., 14. Volginsky village, Petushinski district, 601125. Russia.

Phone: +7 (495) 988-47-94. E-mail: Tyupa@ibcgenerium.ru

² Department of Biotechnology. Faculty of Biotechnology and Industrial Ecology, Mendeleev University of Chemical Technology of Russia. Miusskaya Sq., 9. Moscow 125047. Russia.

Phone: +7 (495) 495-23-79. E-mail: wsezart@yandex.ru

*Supervising author; +Corresponding author

Keywords: cultivation, CHO cells, monoclonal antibodies, CO₂ concentration, perfusion.

Abstract

The concentration of carbon dioxide dissolved in the medium is one of the key components of mammalian cell culture systems. CO₂ is necessary for the growth and cells development, as well as for maintaining the pH level in the nutrient medium. Non-optimal concentrations of carbon dioxide create a serious problem in the cultivation of cell cultures, including Chinese hamster ovary (CHO) cells, which are the main source of recombinant proteins for therapeutic use. The normal physiological concentration of carbon dioxide for most mammalian cells is 4-8%, which corresponds to a partial pressure of carbon dioxide (pCO₂) about 30-55 mm Hg. The deviations from this interval can slow the growth of cell cultures; significantly rearrange their metabolism, reducing the viability and productivity of cells.

In this paper, the effect of extreme concentrations of dissolved carbon dioxide on the growth of CHO cells in both periodic and continuous processes is elucidated.

When cultivating cells in the fed-batch mode, increasing the CO₂ concentration from 5 to 20% significantly inhibits the growth of the culture, while its reduction to 1% alters the metabolism of cells, blocking the phase of lactate utilization. However, it was found that the cell culture is sensitive to CO₂-stress only at an early stage of cultivation and acquires relative stability already in the period of the late exponential phase of growth.

Similar regularities were observed in continuous (perfusion) processes, when in the early stages of culture growth low values of pCO₂ (1%) led to a strong disruption of lactate metabolism. As a result, intensive accumulation of lactic acid in the nutrient medium was observed, as a result of which the viability and productivity of the crop sharply decreased. In the initial period of cultivation, as long as the concentration of cells is not yet so high that they can saturate the environment with autogenic CO₂, it is necessary first of all to organize an uninterrupted supply of carbon dioxide, not allowing it to blow out from the environment, which is relevant not only for industrial, but also for laboratory reactors.

References

- [1] A. Itagaki, G. Kimura. Tes and HEPES buffers in mammalian cell cultures and viral studies: problem of carbon dioxide requirement. *Exp Cell Res.* **1974**. Vol.83. No.2. P.351-361.
- [2] R. Kimura, W.M. Miller. Effects of elevated pCO₂ and/or osmolality on the growth and recombinant tPA production of CHO cells. *Biotechnol Bioeng.* **1996**. Vol.52. P.152-160.
- [3] M.M. Zhu, A. Goyal, D.L. Rank, S.K. Gupta, T.V. Boom, S.S. Lee. Effects of elevated pCO₂ and osmolality on growth of CHO cells and production of antibody-fusion protein B1: a case study. *Biotechnol Prog.* **2005**. Vol.21. P.70-77.
- [4] D.R. Gray, S. Chen, W. Howarth, D. Inlow, B.L. Maiorella. CO₂ in large-scale and high-density CHO cell perfusion culture. *Cytotechnology.* **1996**. Vol.22. P.65-78.
- [5] R. Krapf, C.A. Berry, R.J. Alpern, F.C. Rector. Regulation of cell pH by ambient bicarbonate, carbon dioxide tension and pH in the rabbit proximal convoluted tubule. *J. Clin. Invest.* **1988**. Vol.81. P.381-389.

- [6] M.B. Ganz, G. Boyarski, R.B. Sterzel, W.F. Boron. Arginine vasopressin enhances pHi regulation in the presence of HCO₃⁻ by stimulating three acid-base transport systems. *Nature*. **1989**. Vol.337. P.648-651.
- [7] U. Onken, E. Liefke. Effect of total and partial pressure (oxygen and carbon dioxide) on aerobic microbial processes. *Adv Biochem Eng Biotechnol*. **1989**. Vol.40. P.137-169.
- [8] B. Thorens, P. Vassali. Chloroquine and ammonium chloride prevent terminal glycosylation of immunoglobulins in plasma cells without affecting secretion. *Nature*. **1986**. Vol.321. P.618-620.
- [9] J.E. Oliveira, R. Damiani, K. Vorauer-Uhl, P. Bartolini, M.T. Ribela. Influence of a reduced CO₂ environment on the secretion yield, potency and N-glycan structures of recombinant thyrotropin from CHO cells. *Mol Biotechnol*. **2008**. Vol.39. P.159-166.
- [10] S.S. Mostafa, X. Gu. Strategies for improved dCO₂ removal in large-scale fed-batch cultures. *Biotechnol Prog*. **2003**. Vol.19. No.1. P.45-51.
- [11] C. Harbour, K.S. Low, C.P. Marquis, J.P. Barford. pH control options for hybridoma cultures. *Biotechnol Tech*. **1989**. Vol.3. P.73-78.
- [12] S.K. Yoon, Y. Ahn, K. Han. Enhancement of recombinant erythropoietin production in CHO cells in an incubator without CO₂ addition. *Cytotechnology*. **2001**. Vol.37. P.119-132.
- [13] L.J. Edwards, D. Williams, D. Gardner. Intracellular pH of the mouse preimplantation embryo: amino acids act as buffers of intracellular pH. *Hum Reprod*. **1998**. Vol.13. No.12. P.3441-3448.
- [14] M.S. Kilberg. System A-mediated amino acid transport: metabolic control at the plasma membrane. *Trends Biochem Sci*. **1986**. Vol.11. No.4. P.183-186.
- [15] M.S. Kilberg, B.R. Stevens, D.A. Novak. Recent advances in mammalian amino acid transport. *Annu Rev Nutr*. **1993**. Vol.13. No.1. P.137-165.
- [16] V.M. Zengotita, L.R. Abston, A.E. Schmelzer, W.M. Miller. Selected amino acids protect hybridoma and CHO cells from elevated carbon dioxide and osmolality. *Biotechnol Bioeng*. **2002**. Vol.78. No.7. P.741-752.
- [17] F. Zagari, M. Jordan, M. Stettler, H. Broly, F.M. Wurm. Lactate metabolism shift in CHO cell culture: the role of mitochondrial oxidative activity. *New Biotech*. **2013**. Vol.30. No.2. P.238-245.