

Phosphine oxide as a prospective intermediate of biological processes

© Anton Z. Mindubaev,^{1*} Akosah Yaw Abayie,² and Dmitry G. Yakhvarov¹

¹*Institute of Organic and Physical Chemistry named after A.E. Arbuzov. Kazan Scientific Center of the Russian Academy of Sciences. Arbuzov St., 8. Kazan, 420088. Republic of Tatarstan. Russia.*

E-mail: mindubaev@iopc.ru; mindubaev-az@yandex.ru

²*Kazan (Volga Region) Federal University. University St., 18. Kazan, 420008. Republic of Tatarstan. Russia.*

*Supervising author; ⁺Corresponding author

Keywords: phosphin oxide, phosphine, reduced phosphorus compounds, toxicity, metabolism, lethal metabolites, white phosphorus, biodegradation.

Abstract

In previous works, we demonstrated for the first time the biological conversion of a very hazardous environmental pollutant, an extremely toxic and reactive substance of first class danger – white phosphorus. Although the diversified symptomatology of both acute and chronic poisoning with this substance has been described in detail, the metabolic pathway of white phosphorus has virtually not been studied. In our studies, we sought to fill this gap by establishing the nature of white phosphorus metabolites by NMR and chromatomass spectrometry. Some literary sources have reported on the formation of phosphine in a body, which has been poisoned with elemental phosphorus. There is an assumption that phosphine also exhibits toxic properties not in itself, but through its reactive metabolite, phosphine oxide. In this review, we have made efforts in gathering information on the toxicology of phosphine (and, to a lesser extent, elemental phosphorus) in order to find direct evidence for the formation of phosphine oxide in living cells. Observation on phosphine oxide in a living organism has not been done so far, but there is possibility of its formation as a result of the oxidation of its precursor, phosphine. It is possible that the genotoxicity of white phosphorus, which was discovered in our studies is also due to the formation of phosphine oxide.

References

- [1] F.S. Archibald, I. Fridovich. Oxygen radicals, oxygen toxicity and the life of microorganisms. *Acta medica portuguesa*. **1983**. Vol.4. No.2. P.101-112.
- [2] J.-K. Weng, C. Chapple. The origin and evolution of lignin biosynthesis. *New Phytologist*. **2010**. Vol.187. No.2. P.273-285.
- [3] D. Floudas, M. Binder, R. Riley, K. Barry, R.A. Blanchette, B. Henrissat, A.T. Martínez, R. Otilar, J.W. Spatafora, J.S. Yadav, A. Aerts, I. Benoit, A. Boyd, A. Carlson, A. Copeland, P.M. Coutinho, R.P. de Vries, P. Ferreira, K. Findley, B. Foster, J. Gaskell, D. Glotzer, P. Górecki, J. Heitman, C. Hesse, C. Hori, K. Igarashi, J.A. Jurgens, N. Kallen, P. Kersten, A. Kohler, U. Kües, T.K.A. Kumar, A. Kuo, K. LaButti, L.F. Larrondo, E. Lindquist, A. Ling, V. Lombard, S. Lucas, T. Lundell, R. Martin, D.J. McLaughlin, Ingo Morgenstern, E. Morin, C. Murat, L.G. Nagy, M. Nolan, R.A. Ohm, A. Patyshakuliyeva, A. Rokas, F.J. Ruiz-Dueñas, G. Sabat, A. Salamov, M. Samejima, J. Schmutz, J.C. Slot, F.St. John, J. Stenlid, H. Sun, S. Sun, K. Syed, A. Tsang, A. Wiebenga, D. Young, A. Pisabarro, D.C. Eastwood, F. Martin, D. Cullen, I. V. Grigoriev, D.S. Hibbett. The Paleozoic Origin of Enzymatic Lignin Decomposition Reconstructed from 31 Fungal Genomes. *Science*. **2012**. Vol.336. P.1715-1719.
- [4] S.W. Ragsdale. Life with carbon monoxide. *Crit Rev Biochem Mol Biol*. **2004**. Vol.39. No.3. P.165-195.
- [5] A.Z. Mindubaev, D.G. Yakhvarov. Biodegradation as a method for waste processing: view on the problem. Part 1. The essence of the method. *Butlerov Communications*. **2013**. Vol.33. No.3. P.1-37. ROI: jbc-02/13-33-3-1
- [6] A.Z. Mindubaev, D.G. Yakhvarov. Biodegradation as a method for waste processing: view on the problem. Part 2. Are xenobiotics really xenobiotics? *Butlerov Communications*. **2013**. Vol.34. No.4. P.1-20. ROI: jbc-02/13-34-4-1
- [7] Миндубаев А.З. Биодegradация ксенобиотиков как самозащита природы. *Биомолекула*. **2017**. <https://biomolecula.ru/articles/biodegradatsiia-ksenobiotikov-kak-samozashchita-prirody>

- [8] K. Kawachara, Y. Yakabe, T. Ohide, K. Kida. Evaluation of laboratory – made sludge for an anaerobic biodegradability test and its use for assessment of 13 chemicals. *Chemosphere*. **1999**. Vol.39. No.12. P.2007-2018.
- [9] P.M. Golyshin, H.L. Frederickson, L. Giuliano, R. Rothmel, K.N. Timmis, M.M. Yakimov. Effect of novel biosurfactants on biodegradation of polychlorinated biphenyls by pure and mixed bacterial cultures. *New Microbiol.* **1999**. Vol. 22. No.3. P.257-267.
- [10] S.W. Kang, Y.B. Kim, J.D. Shin, E.K. Kim. Enhanced biodegradation of hydrocarbons in soil by microbial biosurfactant, sophorolipid. *Appl. Biochem. Biotechnol.* **2010**. Vol. 160. No. 3. P. 780-790.
- [11] C.N. Mulligan, N.Y. Raymond, B.F. Gibbs. Heavy metal removal from sediments by biosurfactants. *J. Hazard. Mater.* **2001**. Vol.85. No.1-2. P.112-125.
- [12] T.R. Sandrin, A.M. Chech, R.M. Maier. A rhamnolipid biosurfactant reduces cadmium toxicity during naphthalene biodegradation. *Appl. Environ. Microbiol.* **2000**. Vol.66. No.10. P.4585-4588.
- [13] C. Schippers, K. Gessner, T. Muller, T. Scheper. Microbial degradation of phenantrene by addition of a sophorolipid mixture. *J. Biotechnol.* **2000**. Vol.83. No.3. P.189-198.
- [14] Z. Hua, J. Chen, S. Lun, X. Wang. Influence of biosurfactants produced by *Candida antarctica* on surface properties of microorganism and biodegradation of n-alkanes. *Water Res.* **2003**. Vol.37. No.17. P.4143-4150.
- [15] V.A. Khryachkov, E.A. Saratovskikh, R.N. Yarullin. Study of biodegradation of nitrocellulose using fungus *Fuzarium Solani*. *Chemical Safety*. **2017**. Vol.1. No.1. P.168-176. (russian)
- [16] G.E. Merzlaya, R.A. Afanac'ev. Solution to problem of urban sewage sludge utilization. *Chemical Safety*. **2017**. Vol.1. No.1. P.158-167. (russian)
- [17] T.Z. Esikova, O.V. Volkova, S.A. Taran, A.M. Boronin. Keyrole of the *dcagenesine*-caprolactam catabolism in *Pseudomonas* strains. *Microbiology*. **2015**. Vol.84. No.5. P.726-729.
- [18] S. Saraswat. Patent Analysis on Bioremediation of Environmental Pollutants. *J Bioremed Biodeg.* **2014**. Vol.5. No.6. 251p.
- [19] A.E. Kuznetsov, N.B. Gradova, S.V. Lushnikov, M. Engelhart, T. Weisser, M.V. Chebotareva. Applied ecobiotechnology. *Textbook for high school. Ed. BINOM*. 2nd ed. **2015**. Vol.1. P.629. (russian)
- [20] C. Holliger, G. Schraa, A.J.M. Stams, A. J.B. Zehnder. Reductive dechlorination of 1,2-dichloroethane and chloroethane by cell suspensions of methanogenic bacteria. *Biodegradation*. **1990**. Vol.1. No.4. P.253-261.
- [21] J.A. Dijk, J. Gerritse, G. Schraa, A.J.M. Stams. Degradation pathway of 2-chloroethanol in *Pseudomonas stutzeri* strain JJ under denitrifying conditions. *Arch Microbiol.* **2004**. Vol.182. No.6. P.514-519.
- [22] E.K. Vasileva, V.N. Beschkov. Review on biodegradation of 1,2-dichloroethane. *Scientific works of UFT "Food science, engineering and technologies"*. **2012**. Vol. lix- 2012. P.669-674.
- [23] L.A. Vanderberg, B.L. Burbach, J.J. Perry. Biodegradation of trichloroethylene by *Mycobacterium vaccae*. *Can J Microbiol.* **1995**. Vol.41. No.3. P.298-301.
- [24] T. Bosma, J. Damborský, G. Stucki, D.B. Janssen. Biodegradation of 1,2,3-Trichloropropane through Directed Evolution and Heterologous Expression of a Haloalkane Dehalogenase Gene. *Applied and environmental microbiology*. **2002**. Vol.68. No.7. P.3582-3587.
- [25] T. Toraya, T. Oka, M. Ando, M. Yamanishi, H. Nishihara. Novel Pathway for Utilization of Cyclopropanecarboxylate by *Rhodococcus rhodochrous*. *Applied and environmental microbiology*. **2004**. Vol.70. No.1. P.224-228.
- [26] J.G. Schiller, A.E. Chung. The Metabolism of Cyclopropanecarboxylic Acid. *The Journal of Biological Chemistry*. **1970**. Vol.245. No.21. P.5857-5864.
- [27] W.J.J. Van den Tweel, J.A.M. De Bont. Metabolism of 3-Butyn-1-ol by *Pseudomonas* BB1. *Journal of General Microbiology*. **1985**. Vol.131. P.3155-3162.
- [28] S. Hartmans, M.W. Jansen, M.J. Van Der Werf, J.A.M. De Bont. Bacterial metabolism of 3-chloroacrylic acid. *Journal of General Microbiology*. **1991**. Vol.137. P.2025-2032.
- [29] R. Gurusamy, S. Natarajan. Current Status on Biochemistry and Molecular Biology of Microbial Degradation of Nicotine. *The ScientificWorld Journal*. **2013**. Vol.2013. Article ID 125385. P.1-15.
- [30] M. Mihasan, C.-B. Chiribau, T. Friedrich, V. Artenie, R. Brandsch. An NAD(P)H-Nicotine Blue Oxidoreductase Is Part of the Nicotine Regulon and May Protect *Arthrobacter nicotinovorans* from Oxidative Stress during Nicotine Catabolism. *Applied and environmental microbiology*. **2007**. Vol.73. No.8. P.2479-2485.
- [31] D.R. Kadavy, B.A. Plantz, C.A. Shaw, J. Myatt, T.A. Kokjohn, K. Nickerson. Microbiology of the Oil Fly, *Helaeomyia petrolei*. *Applied and environmental microbiology*. **1999**. Vol.65. No.4. P.1477-1482.
- [32] J.R. Lloyd. Microbial reduction of metals and radionuclides. *FEMS Microbiology Reviews*. **2003**. Vol.27. No.2-3. P.411-425.

- [33] S. Yoshida, K. Hiraga, T. Takehana, I. Taniguchi, H. Yamaji, Y. Maeda, K. Toyohara, K. Miyamoto, Y. Kimura, K. Oda. A bacterium that degrades and assimilates poly(ethylene terephthalate). *Science*. **2016**. Vol.351. No.6278. P.1196-1199.
- [34] V.B. Pinheiro, A.I. Taylor, C. Cozens, M. Abramov, M. Renders, S. Zhang, J.C. Chaput, J. Wengel, S.-Y. Peak-Chew, S.H. McLaughlin, P. Herdewijn, P. Holliger. Synthetic Genetic Polymers Capable of Heredity and Evolution. *Science*. **2012**. Vol.336. No.6079. P.341-344.
- [35] G.F. Joyce. Toward an Alternative Biology. *Science*. **2012**. Vol.336. No.6079. P.307-308.
- [36] D.E. Mills, W.D. Baugh, H.A. Conner. Studies on the Formation of Acrolein in Distillery Mash. *Appl Microbiol*. **1954**. Vol.2. No.1. P.9-13.
- [37] K.C. Nicolaou, A.L. Smith, E.W. Yue. Chemistry and biology of natural and designed enediynes. *Proc. Natl. Acad. Sci. USA*. **1993**. Vol.90. No.13. P.5881-5888.
- [38] P. Jumaryatno, K. Rands-Trevor, J.T. Blanchfield, M.J. Garson. Isocyanates in marine sponges: Axisocyanate-3, a new sesquiterpene from *Acanthella cavernosa*. *ARKIVOC*. **2007**. Vol.2007. No.7. P.157-166.
- [39] G.D. Peiser, T.-T. Wang, N.E. Hoffman, S.F. Yang, H.-W. Liut, C.T. Walsh. Formation of cyanide from carbon 1 of 1-aminocyclopropane-1-carboxylic acid during its conversion to ethylene. *Proc. Natl. Acad. Sci. USA*. **1984**. Vol.81. No.10. P.3059-3063.
- [40] B. Van de Poel, D. Van Der Straeten. 1-Aminocyclopropane-1-carboxylic acid (ACC) in plants: more than just the precursor of ethylene! *Frontiers in Plant Science. Plant Physiology*. **2014**. Vol.5. P.1-11.
- [41] R. Deml, K. Dettner. Comparative morphology and secretion chemistry of the scoli in caterpillars of *Hyalophora cecropia*. *Naturwissenschaften*. **2003**. Vol.90. No.10. P.460-463.
- [42] M. Saarivirta, A.I. Virtanen. A Method for Estimating Benzyl Isothiocyanate, Benzyl Thiocyanate, and Benzyl Nitrile in the Crushed, Moistened Seeds of *Lepidium sativum*. *Acta Chemica Scandinavica*. **1964**. Vol.17. P.74-78.
- [43] K.-H. van Pée. Biosynthesis of halogenated metabolites by bacteria. *Annu. Rev. Microbiol*. **1996**. Vol.50. P.375-399.
- [44] J. Fuska, M. Šturdíková, B. Proksa. Biotransformation of benzyl thiocyanate by *Streptomyces aureofaciens*. *Letters in Applied Microbiology*. **1994**. Vol.19. No.3. P.124-125.
- [45] L.A.D. Williams, H. Rosner, H.G. Levy, E.N. Barton. A Critical Review of the Therapeutic Potential of Dibenzyl Trisulphide Isolated from *Petiveria alliacea* L (Guinea hen weed, anamu). *West Indian Med J*. **2007**. Vol.56. No.1. P.17-21.
- [46] J. Zi, S. Li, M. Liu, M. Gan, S. Lin, W. Song, Y. Zhang, X. Fan, Y. Yang, J. Zhang, J. Shi, D. Di. Glycosidic Constituents of the Tubers of *Gymnadenia conopsea*. *J. Nat. Prod*. **2008**. Vol.71. No.5. P.799-805.
- [47] G. Preti, E. Thaler, C.W. Hanson, M. Troy, J. Eades, A. Gelperin. Volatile compounds characteristic of sinus-related bacteria and infected sinus mucus: Analysis by solid-phase microextraction and gas chromatography–mass spectrometry. *Journal of Chromatography B*. **2009**. Vol.877. No.22. P.2011-2018.
- [48] T. Yoshioka, J.A. Krauser, F.P. Guengerich. Microsomal Oxidation of Tribromoethylene and Reactions of Tribromoethylene Oxide. *Chem. Res. Toxicol*. **2002**. Vol.15. No.11. P.1414-1420.
- [49] E.A. Lock, C.J. Reed. Trichloroethylene: Mechanisms of Renal Toxicity and Renal Cancer and Relevance to Risk Assessment. *Toxicological Sciences*. **2006**. Vol.91. No.2. P.313-331.
- [50] L.H. Lash, J.W. Fisher, J.C. Lipscomb, J.C. Parker. Metabolism of Trichloroethylene. *Environmental Health Perspectives*. **2000**. Vol.108. P.177-200.
- [51] G. Segev, E. Yas-Natan, A. Shlosberg, I. Aroch. Alpha-chloralose poisoning in dogs and cats: A retrospective study of 33 canine and 13 feline confirmed cases. *The Veterinary Journal*. **2006**. Vol.172. No.1. P.109-113.
- [52] D.M.E. Szebenyi, F.N. Musayev, M.L. di Salvo, M.K. Safo, V. Schirch. Serine Hydroxymethyltransferase: Role of Glu75 and Evidence that Serine Is Cleaved by a Retroaldol Mechanism. *Biochemistry*. **2004**. Vol.43. No.22. P.6865-6876.
- [53] M.L. di Salvo, S.G. Remesh, M. Vivoli, M.S. Ghatge, A. Paiardini, S. D'Aguanno, M.K. Safo, R. Contestabile. On the catalytic mechanism and stereospecificity of *Escherichia coli* L-threonine aldolase. *FEBS Journal*. **2014**. Vol.281. No.1. P.129-145.
- [54] C.D. Lyons, S. Katz, R. Bartha. Mechanisms and Pathways of Aniline Elimination from Aquatic Environments. *Appl. Environ. Microbiol*. **1984**. Vol.48. No.3. P.491-496.
- [55] L.M. Blair, J. Sperry. Natural Products Containing a Nitrogen–Nitrogen Bond. *J. Nat. Prod*. **2013**. Vol.76. No.4. P.794-812.
- [56] J. Shi, W. Lu, F. Sun. Benzoic acid-enriched plant extracts and use thereof. *Patent US 2013/0296432 A1. Pub. Date: Nov. 7, 2013*.

- [57] T. Gulder, P.S. Baran. Strained cyclophane natural products: Macrocyclization at its limits. *Nat. Prod. Rep.* **2012**. Vol.29. No.8. P.899-934.
- [58] D.L. Kaplan, J.H. Cornell, A.M. Kaplan. Decomposition of Nitroguanidine *Environ. Sci. Technol.* **1982**. Vol.16. No.8. P.488-492.
- [59] T. Kamo, M. Endo, M. Sato, R. Kasahara, H. Yamaya, S. Hiradate, Y. Fujii, N. Hirai, M. Hirota. Limited distribution of natural cyanamide in higher plants: Occurrence in *Vicia villosa* subsp. *varia*, *V. cracca*, and *Robinia pseudo-acacia*. *Phytochemistry*. **2008**. Vol.69. No.5. P.1166-1172.
- [60] N. Belay, L. Daniels. Production of Ethane, Ethylene, and Acetylene from Halogenated Hydrocarbons by Methanogenic Bacteria. *Applied and environmental microbiology*. **1987**. Vol.53. No.7. P.1604-1610.
- [61] C. Paul, G. Pohnert. Production and role of volatile halogenated compounds from marine algae. *Nat. Prod. Rep.* **2011**. Vol.28. No.186. P.186-195.
- [62] M.S.P. Mtolera, J. Collén 2, M. Pedersén, A. Ekdahl, K. Abrahamsson, A.K. Semesi. Stress-induced production of volatile halogenated organic compounds in *Eucheuma denticulatum* (Rhodophyta) caused by elevated pH and high light intensities. *Eur. J. Phycol.* **1996**. Vol.31. P.89-95.
- [63] T. Jojima, M. Inui, H. Yukawa. Production of isopropanol by metabolically engineered *Escherichia coli*. *Appl Microbiol Biotechnol.* **2008**. Vol.77. No.6. P.1219-1224.
- [64] R. Jain, Y. Yan. Dehydratase mediated 1-propanol production in metabolically engineered *Escherichia coli*. *Microbial Cell Factories*. **2011**. Vol.10:97. P.1-10.
- [65] E. Rentschler, K. Schuh, M. Krewinkel, C. Baur, W. Claaßen, S. Meyer, B. Kuschel, T. Stressler, L. Fischer. Enzymatic production of lactulose and epilactose in milk. *J. Dairy Sci.* **2015**. Vol.98. No.10. P.6767-6775.
- [66] T.B. Taylor, G. Mulley, A.H. Dills, A.S. Alsohim, L.J. McGuffin, D.J. Studholme, M.W. Silby, M.A. Brockhurst, L.J. Johnson, R.W. Jackson. Evolutionary resurrection of flagellar motility via rewiring of the nitrogen regulation system. *Science*. **2015**. Vol.347. No.6225. P.1014-1017.
- [67] A.Z. Mindubaev, D.G. Yakhvarov. Phosphorus: properties and application. *Butlerov Communications*. **2014**. Vol.39. No.7. P.1-24. ROI: jbc-02/14-39-7-1
- [68] H.-G. Hoppe. Phosphatase activity in the sea. *Hydrobiologia*. **2003**. Vol.493. No.1-3. P.187-200.
- [69] C.T. Reinhard, N.J. Planavsky, B.C. Gill, K. Ozaki, L.J. Robbins, T.W. Lyons6, W.W. Fischer, C. Wang, D.B. Cole, K.O. Konhauser. Evolution of the global phosphorus cycle. *Nature*. **2016**. doi: 10.1038/nature20772.
- [70] T.K. Tromp, P. VanCapellen, R.M. Key. A global model for early diagenesis of organic carbon and organic phosphorus in marine sediments. *Geochimica et Cosmochimica Acta*. **1995**. Vol.59. No.7. P.1259-1284.
- [71] R.J. Seviour, T. Mino, M. Onuki. The microbiology of biological phosphorus removal in activated sludge systems. *FEMS Microbiology Reviews*. **2003**. Vol.27. No.1. P.99-127.
- [72] L. D'Croze, J.B. Del Rosario, J.A. Gómez. Upwelling and phytoplankton in the Bay of Panama. **1991**. *Rev. Biol. Trop.* Vol.39. No.2. P.233-241.
- [73] A.D. Cembella, N.J. Antia, P. J. Harrison. The utilization of inorganic and organic phosphorous compounds as nutrients by eukaryotic microalgae: a multidisciplinary perspective: part 1. *CRC Critical Reviews in Microbiology*. **1984**. Vol.10. No.4. P.317-391.
- [74] A.D. Cembella, N.J. Antia, P. J. Harrison. The utilization of inorganic and organic phosphorous compounds as nutrients by eukaryotic microalgae: a multidisciplinary perspective: part 2. *CRC Critical Reviews in Microbiology*. **1984**. Vol.11. No.1. P.13-81.
- [75] G. Hu, K.S. Lim, N. Horvitz, S.J. Clark, D. R. Reynolds, N. Sapir, J.W. Chapman. Mass seasonal bioflows of high-flying insect migrants. *Science*. **2016**. Vol.354. No.6319. P.1584-1587.
- [76] J.B. León, C.M. Sullivan, A. R. Sehgal. The Prevalence of Phosphorus-Containing Food Additives in Top-Selling Foods in Grocery Stores. *Journal of Renal Nutrition*. **2013**. Vol.23. No.4. P.265-270.
- [77] H. Rodríguez, R. Fraga. Phosphate solubilizing bacteria and their role in plant growth promotion. *Biotechnology Advances*. **1999**. Vol.17. No.4-5. P.319-339.
- [78] P. Gyaneshwar, G.N. Kumar, L.J. Parekh, P.S. Poole. Role of soil microorganisms in improving P nutrition of plants. *Plant and Soil*. **2002**. Vol.245. No.1. P.83-93.
- [79] R.A. Moen, J. Pastor, Y. Cohen. Antler growth and extinction of Irish elk. *Evolutionary Ecology Research*. **1999**. Vol.1. P.235-249.
- [80] J. Tissier, J.-C. Rage, M. Laurin. Exceptional soft tissues preservation in a mummified frog-eating Eocene Salamander. *Peer J*. **2017**. Vol.5. No.3861. P.1-14.
- [81] G.E. Guskov, I.S. Vasilchenko. Antimony and bismuth as a possible basis of life. *Scientific thought of Caucasus*. **2012**. No.3 (71). P.94-97.

- [82] A.Z. Mindubaev, Y.A. Akosah, F.K. Alimova, D.M. Afordoanyi, R.M. Kagirov, S.T. Minzanova, L.G. Mironova, O.G. Sinyashin, D.G. Yakhvarov. On the White Phosphorus Degradation by Wastewater Mud. *Uchenye Zapiski Kazanskogo Universiteta. Seriya Estestvennyye Nauki*. **2011**. Vol.153. No.2. P.110-119.
- [83] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, C. Bolormaa, A.D. Voloshina, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. The possibility for anaerobic detoxication of white phosphorus. *Butlerov Communications*. **2013**. Vol.33. No.1. P.22-34. ROI: jbc-02/13-33-1-22
- [84] A.Z. Mindubaev, A.D. Voloshina, D.G. Yakhvarov. Biological degradation of white phosphorus: feasibility and prospects. *Butlerov Communications*. **2013**. Vol.33. No.2. P.1-17. ROI: jbc-02/13-33-2-1
- [85] A.Z. Mindubaev, A.D. Voloshina, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov, F.K. Alimova, S.C. Ahossiyenagbe, A.C. Bolormaa. Possibility of anaerobic biodegradation of white phosphorus. *The North Caucasus Ecological Herald*. **2013**. Vol.9. No.2. P.4-15.
- [86] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. New confirmation for white phosphorus biodegradation. *Butlerov Communications*. **2013**. Vol.36. No.10. P.1-12. ROI: jbc-02/13-36-10-1
- [87] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, C. Bolormaa, A.D. Voloshina, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Microbial metabolism of the white phosphorus. *Butlerov Communications*. **2013**. Vol.36. No.12. P.34-52. ROI: jbc-02/13-36-12-34
- [88] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Relationship between microbial metabolism and rate of destruction of white phosphorus in sewage sludge. *The North Caucasus Ecological Herald*. **2014**. Vol.10. No.1. P.88-96.
- [89] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Metabolites and tolerant microflora in substrates with white phosphorus 0.1%. *Butlerov Communications*. **2014**. Vol.37. No.3. P.67-78. ROI: jbc-02/14-37-3-67
- [90] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, E.V. Gorbachuk, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Metabolic pathway of white phosphorus. *The North Caucasus Ecological Herald*. **2014**. Vol.10. No.3. P.36-46. (russian)
- [91] C. Bolormaa, K.A. Saparmyradov, F.K. Alimova, A.Z. Mindubaev. Comparison of phytotoxicity indices, fungicidal and bactericidal activity of *Streptomyces* from different habitats. *Butlerov Communications*. **2014**. Vol.38. No.6. P.147-152. ROI: jbc-02/14-38-6-147
- [92] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, C. Bolormaa, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, L.G. Mironova, A.V. Pankova, D.G. Yakhvarov. Decontamination of industrial waste water containing white phosphorus, using SALT microflora. *Ecology and Safety Journal*. **2014**. No.1-2. P.68-72. (russian)
- [93] A.Z. Mindubaev, F.K. Alimova, S.C. Ahossiyenagbe, E.V. Gorbachuk, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Resistant microflora in substrata with phosphorus content 0.1% by mass, and its cultivation in artificial media. *The North Caucasus Ecological Herald*. **2014**. Vol.10. No.4. P.66-74. (russian)
- [94] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, S.C. Ahossiyenagbe, F.K. Alimova, S.T. Minzanova, L.G. Mironova, A.V. Pankova, C. Bolormaa, K.A. Saparmyradov, D.G. Yakhvarov. White phosphorus as a new object of biological destruction. *Butlerov Communications*. **2014**. Vol.40. No.12. P.1-26. ROI: jbc-02/14-40-12-1
- [95] A.Z. Mindubaev, F.K. Alimova, D.G. Yakhvarov, C. Bolormaa, K.A. Saparmyradov. Comparison of phytotoxicity, fungicidal and bactericidal activity of *Streptomyces* from different biotopes. Determination of species rank for strain A8. *The North Caucasus Ecological Herald*. **2015**. Vol.11. No.1. P.51-58. (russian)
- [96] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, F.K. Alimova, S.T. Minzanova, L.G. Mironova, K.A. Saparmyradov, K.R. Khayarov, D.G. Yakhvarov. The inclusion white phosphorus in the natural cycle of matter. Cultivation of resistant microorganisms. *Butlerov Communications*. **2015**. Vol.41. No.3. P. 54-81. ROI: jbc-02/15-41-3-54
- [97] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, S.T. Minzanova, L.G. Mironova, F.K. Alimova, K.A. Saparmyradov, D.G. Yakhvarov. Increase of resistance to white phosphorus in microorganisms as a result of directed selection: biochemical analysis of *streptomyces* sp. A8 strain. *The North Caucasus Ecological Herald*. **2015**. Vol.11. No.3. P.10-18. (russian)
- [98] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, F.K. Alimova, K.A. Saparmyradov, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Ecotoxicant white phosphorus as phosphoric fertilizer for microorganisms. *Journal of Ecology and Industrial Safety*. **2015**. No.1-2. P.46-51. (russian)

- [99] A.Z. Mindubaev, S.T. Minzanova, L.G. Mironova, F.K. Alimova, D.E. Belostotskiy, D.G. Yakhvarov. The effect of the amaranth phytomass on the white phosphorus biodegradation rate. *The North Caucasus Ecological Herald*. **2015**. Vol.11. No.4. P.73-79. (russian)
- [100] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, S. Z. Validov, N.V. Kulik, F.K. Alimova, S.T. Minzanova, L.G. Mironova, D.E. Belostotskiy, K.A. Saparmyradov, R.I. Tukhbatova, D.G. Yakhvarov. Adaptation of microorganisms to white phosphorus as a result of directed selection. Genetic identification of sustainable *Aspergillus* and metabolic profiling of *Streptomyces* A8. *Butlerov Communications*. **2015**. Vol.44. No.12. P.1-28. ROI: jbc-02/15-44-12-1
- [101] A.Z. Mindubaev, S.Z. Validov, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, F.K. Alimova, S.T. Minzanova, R.I. Tukhbatova, L.G. Mironova, D.G. Yakhvarov. Identification of resistant to white phosphorus *Aspergillus*. *The North Caucasus Ecological Herald*. **2016**. Vol.12. No.1. P.70-75. (russian).
- [102] A.Z. Mindubaev. Biodegradation of white phosphorus: how a poison became a fertilizer. *Biomolecula*. **2016**. <http://biomolecula.ru/content/1932>
- [103] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, S.T. Minzanova, L.G. Mironova, F.K. Alimova, D.G. Yakhvarov. The detoxication of white phosphorus containing wastewaters, by microflora. *Russian Journal of Applied Ecology*. **2015**. No.3. P.42-47. (russian)
- [104] A.Z. Mindubaev, A.D. Voloshina, S. Z. Validov, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov, A.Y. Akkizov. *Aspergillus niger* AM1 culture growth in medium with two phosphorus sources. The validity of the definition "biodegradation" with respect to white phosphorus. *Butlerov Communications*. **2016**. Vol.46. No.5. P.1-20. ROI: jbc-02/16-46-5-1
- [105] A.Z. Mindubaev, A.D. Voloshina, S.T. Minzanova. Growth of bacterial culture in a medium with potassium phosphite as a sole source of phosphorus. *The North Caucasus Ecological Herald*. **2016**. Vol.12. No.3. P.81-84. (russian)
- [106] A.Z. Mindubaev, K.A. Saparmyradov, E.V. Gorbachuk, A.V. Pankova. Selection of microorganisms for resistance to white phosphorus. *Russian Journal of Applied Ecology*. **2016**. No.2. P.42-46. (russian)
- [107] A.Z. Mindubaev, F.K. Alimova, A.D. Voloshina, E.V. Gorbachuk, N.V. Kulik, S.T. Minzanova, R.I. Tukhbatova, D.G. Yakhvarov. Method for detoxification of white phosphorus using microorganism strain *Trichoderma asperellum* VKPM F-1087. *Patent RF No 2603259 from 1.11.2016. Bul. 33. Reciprocity date 28. 07. 2015. Registration number 2015131380 (048333). The decision to grant a patent on 29. 08. 2016*. (russian)
- [108] A.Z. Mindubaev, E.V. Babynin, A.D. Voloshina, S.Z. Validov, N.V. Kulik, S.T. Minzanova, L.G. Mironova, A.Y. Akkizov, D.G. Yakhvarov. Evaluation of white phosphorus genotoxicity. Growth of bacterial culture in a medium with potassium phosphite as a sole source of phosphorus. *Butlerov Communications*. **2016**. Vol.47. No.7. P.1-20. ROI: jbc-02/16-47-7-1
- [109] A.Z. Mindubaev, A.D. Voloshina, N.V. Kulik, Sh. Z. Validov, E.V. Babynin, K.A. Saparmyradov, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Selection of cultures of microorganisms for resistance to white phosphorus, and their identification. *Journal of Ecology and Industrial Safety*. **2016**. No.2. P.22-27. (russian)
- [110] A.Z. Mindubaev, K.A. Saparmyradov, F.K. Alimova. Comparison of antagonistic properties of *Streptomyces* from different biotopes. *Russian Journal of Applied Ecology*. **2016**. No.3. P.28-32. (russian)
- [111] A.Z. Mindubaev, A.D. Voloshina, S.T. Minzanova. Possibility of decontamination of decontamination by using white phosphorus. *The North Caucasus Ecological Herald*. **2016**. Vol.12. No.4. P.63-70. (russian)
- [112] A.Z. Mindubaev, E.V. Babynin, A.D. Voloshina, I.F. Sakhapov, N.V. Kulik, S.Z. Validov, S.T. Minzanova, L.G. Mironova, A.Y. Akkizov, D.G. Yakhvarov. Genotoxicity of white phosphorus. *Butlerov Communications*. **2017**. Vol.49. No.1. P.1-20. ROI: jbc-02/17-49-1-1
- [113] A.Z. Mindubaev, A.D. Voloshina, S.Z. Validov. *Aspergillus niger* AM1 culture growth in medium with two phosphorus sources. effectiveness of white phosphorus sterilization with acetone. *The North Caucasus Ecological Herald*. **2017**. Vol.13. No.1. P.47-54. (russian)
- [114] A.Z. Mindubaev, A.D. Voloshina, S. Z. Validov, N.V. Kulik, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov, A.Y. Akkizov. *Aspergillus niger* AM1 culture growth in medium with two phosphorus sources. The validity of the definition "biodegradation" with respect to white phosphorus. *Butlerov Communications*. **2016**. Vol.46. No.5. P.1-20. ROI: jbc-02/16-46-5-1
- [115] A.Z. Mindubaev. Biodegradation of white phosphorus: as a poison became a fertilizer. *A.E. Arbuzova's IOPC. 2016. Yearbook. Kazan, KFTI, KazNC of the Russian Academy of Sciences*. **2017**. P.93-105. (russian)

- [116] A.Z. Mindubaev, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. The effect of the amaranth green mass on the white phosphorus degradation rate. *Russian Journal of Applied Ecology*. **2017**. No.1. P.50-54. (russian)
- [117] A.Z. Mindubaev AZ, A.D. Voloshina, S.Z. Validov, D.G. Yakhvarov. Biodegradation of white phosphorus. *Nature*. **2017**. No.5. P.29-43. (russian)
- [118] A.Z. Mindubaev, Sh.Z. Validov, E.V. Babynin. Research of white phosphorus genotoxicity. *The North Caucasus Ecological Herald*. **2017**. Vol.13. No.2. P. 38-44.
- [119] A.Z. Mindubaev, A.D. Voloshina, N.V. Kulik, K.A. Saparmyradov, Kh.R. Khayarov, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Anaerobic detoxication of white phosphorus by sewage sludge microorganisms. *Chemical safety*. **2017**. Vol.1. No.1. P.177-192. (russian)
- [120] A.Z. Mindubaev, A.D. Voloshina, E.V. Gorbachuk, S.Z. Validov, N.V. Kulik, F.K. Alimova, S.T. Minzanova, L.G. Mironova, D.E. Belostotsky, K.A. Saparmyradov, R.I. Tukhbatova, D.G. Yakhvarov. Adaptation of microorganisms to white phosphorus as a result of directed selection. Genetic identification of sustainable *Aspergillus* and metabolic profiling of *Streptomyces* A8. *Butlerov Communications*. **2015**. Vol.44. No.12. P.1-28. ROI: jbc-02/15-44-12-1
- [121] A.Z. Mindubaev, E.V. Babynin, S.Z. Validov, A.D. Voloshina, N.V. Kulik, S.T. Minzanova, L.G. Mironova, A.Y. Akkizov, D.G. Yakhvarov. Evaluation of white phosphorus genotoxicity. Growth of bacterial culture in a medium with potassium phosphite as a sole source of phosphorus. *Butlerov Communications*. **2016**. Vol.47. No.7. P.1-20. ROI: jbc-02/16-47-7-1
- [122] A.Z. Mindubaev, E.V. Babynin, A.D. Voloshina, I.F. Sakhapov, N.V. Kulik, S.Z. Validov, S.T. Minzanova, L.G. Mironova, A.Y. Akkizov, D.G. Yakhvarov. Genotoxicity of white phosphorus. *Butlerov Communications*. **2017**. Vol.49. No.1. P.1-20. ROI: jbc-02/17-49-1-1
- [123] A.Z. Mindubaev, A.D. Voloshina, D.G. Yakhvarov. Biological degradation of white phosphorus: feasibility and prospects. *Butlerov Communications*. **2013**. Vol.33. No.2. P.1-17. ROI: jbc-02/13-33-2-1
- [124] A.Z. Mindubaev, E.V. Babynin, A.D. Voloshina, I.F. Sakhapov, D.G. Yakhvarov. Study of the white phosphorus genotoxicity. *Russian Journal of Applied Ecology*. **2017**. No.2. P.42-46.
- [125] A.Z. Mindubaev, A.D. Voloshina, Kh.R. Khayarov, I.F. Sakhapov, E.K. Badeeva, A.S. Strobykina, Sh.Z. Validov, V.M. Babaev, S.T. Minzanova, L.G. Mironova, A.Y. Abaye, D.G. Yakhvarov. Dynamics of white phosphorus transformation by a culture of black aspergill. *Butlerov Communications*. **2017**. Vol.51. No.8. P.1-26. ROI: jbc-02/17-51-8-1
- [126] A.Z. Mindubaev, E.V. Babynin, A.D. Voloshina. Estimation of mutagenic and antimutagenic properties of white phosphorus using SOS-lux test. *The North Caucasus Ecological Herald*. **2017**. Vol.13. No.4. P.40-45. (russian)
- [127] A.Z. Mindubaev, A.D. Voloshina, E.V. Babynin, E.K. Badeeva, Kh.R. Khayarov, S. T. Minzanova, D.G. Yakhvarov. Microbiological degradation of white phosphorus. *Ecology and Industry of Russia*. **2018**. Vol.22. No.1. P.33-37. (russian)
- [128] A.Z. Mindubaev, A.D. Voloshina, N.V. Kulik, K.A. Saparmyradov, Kh.R. Khayarov, S.T. Minzanova, L.G. Mironova, D.G. Yakhvarov. Involving white phosphorus into natural cycle of biogenic elements. *Chemical safety*. **2017**. Vol.1. No.2. P.205-220. (russian)
- [129] A.Z. Mindubaev, A.D. Voloshina, E.V. Babynin, Sh.Z. Validov, K.A. Saparmyradov, Kh.R. Khayarov, E.K. Badeeva, S.T. Minzanova, L.G. Mironova, Y.A. Akosah, D.G. Yakhvarov. Neutralization of white phosphorus by means of microbiological decomposition. *Butlerov Communications*. **2017**. Vol.52. No.12. P.87-118. ROI: jbc-02/17-52-12-87
- [130] S.H. Han, Y.H. Zhuang, H.X. Zhang, Z.J. Wang, J.Z. Yang. Phosphine and methane generation by the addition of organic compounds containing carbon-phosphorus bonds into incubated Soil. *Chemosphere*. **2002**. Vol.49. No.6. P.651-657.
- [131] R.O. Jenkins, A. Morris, P.J. Craig, A.W. Ritchie, N. Ostah. Phosphine generation by mixed – and monoseptic – cultures of anaerobic bacteria. *The Science of the Total Environment*. **2000**. Vol.250. No.1-3. P.73-81.
- [132] J. Geng, X. Jin, Q. Wang, X. Niu, X. Wang, M. Edwards, D. Glindemann. Matrix bound phosphine formation and depletion in eutrophic lake sediment fermentation-simulation of different environmental factors. *Anaerobe*. **2005**. Vol.11. Iss.8. P.273-279.
- [133] C. Haifeng, L. Ji'ang, Z. Yahui, D. Glindemann. Emission sources of atmospheric phosphine and simulation of phosphine formation. *Science in China (Series B)*. **2000**. Vol.43. No.2. P.162-168.
- [134] D. Glindemann, M. Edwards, O. Schrems. Phosphine and methylphosphine production by simulated lightning – a study for the volatile phosphorus cycle and cloud formation in the earth atmosphere. *Atmospheric Environment*. **2004**. Vol.38. No.39. P.6867-6874.

- [135] L. Zhipei, J. Shengfen, W. Baojun, Z. Tao, L. Shuangjiang. Preliminary investigation on the role of microorganisms in the production of phosphine. *Journal of Environmental Sciences*. **2008**. Vol.20. No.7. P.885-890.
- [136] G. Gassmann, D. Glindemann. Phosphine (PH₃) in the biosphere. *Angewandte Chemie, International Edition English*. **1993**. Vol.32. No.5. P.761-763.
- [137] D. Glindemann, U. Stottmeister, A. Bergmann. Free phosphine from the anaerobic biosphere. *Environmental Science and Pollution Research*. **1996**. Vol.3. No.1. P.17-19.
- [138] J.H. Weber. Volatile hydride and methyl compounds of selected elements formed in the marine environment. *Marine Chemistry*. **1999**. Vol.65. No.1-2. P.67-75.
- [139] M.A. Pasek, J.M. Sampson, Z. Atlas. Redox chemistry in the phosphorus biogeochemical cycle. *PNAS*. **2014**. Vol.111. No.43. P.15468-1547.
- [140] U. Deppenmeier, V. Müller. Life close to the thermodynamic limit: how methanogenic archaea conserve energy. *Results Probl. Cell Differ*. **2008**. Vol.45. P.123-152.
- [141] J.W. Lengeler, G. Drews, H.G. Schlegel. Modern Microbiology. Prokaryotes. Vol.2. Moscow: «Mir». **2005**. P.493. (russian)
- [142] P.A. Moore, K.R. Reddy. Role of Eh and pH on Phosphorus Geochemistry in Sediments of Lake Okeechobee, Florida. *Journal of Environmental Quality*. **1994**. Vol.23. No.5. P.955-964.
- [143] O.F.X. Donard, J.H. Weber. Volatilization of tin as stannane in anoxic environments. *Nature*. **1988**. Vol.332. No.6162. P.339-341.
- [144] D.E. Bryant, D. Greenfield, R.D. Walshaw, B.R.G. Johnson, B. Herschy, C. Smith, M.A. Pasek, R. Telford, I. Scowen, T. Munshi, H.G.M. Edwards, C.R. Cousins, I.A. Crawford, T.P. Kee. Hydrothermal modification of the Sikhote-Alin iron meteorite under low pH geothermal environments. A plausibly prebiotic route to activated phosphorus on the early Earth. *Geochimica et Cosmochimica Acta*. **2013**. Vol.109. P.90-112.
- [145] R.M. Kagiroy, A.V. Voloshin, M.K. Kadirov, I.R. Nizameev, O.G. Sinyashin, D.G. Yakhvarov. Selective synthesis of nanosized palladium phosphides from white phosphorus. *Mendeleev Commun*. **2011**. Vol.21. No.4. P.1-4.
- [146] S. Carencio, M. Demange, J. Shi, C. Boissière, C. Sanchez, P. Le Flochz, N. Mézailles. White phosphorus and metal nanoparticles: a versatile route to metal phosphide nanoparticles. *Chem. Commun*. **2010**. Vol.46. No.30. P.5578-5580.
- [147] K. Adolfsson, M. Schneider, G. Hammarin, U. Häcker, C.N. Prinz. Ingestion of gallium phosphide nanowires has no adverse effect on *Drosophila* tissue function. *Nanotechnology*. **2013**. Vol.24. No.28. P.1-7.
- [148] R. Ripan, I. Chetyanu. Inorganic chemistry. Chemistry of Metals. Moscow: Mir. **1971**. Vol.1. P.561. (russian)
- [149] G.L. Long, C.B. Boss. Removal of phosphine from acetylene. *Anal. Chem*. **1981**. Vol.53. No.14. P.2363-2365.
- [150] I.S. Nikandrov, S.I. Smirnov. Kinetics of oxidation of phosphorus with water. *Journal of Applied Chemistry*. **1983**. Vol.56. No.4. P.883-885. (russian)
- [151] V.M. Vorotyntsev, G.M. Mochalov, S.S. Balabanov. Synthesis of phosphine by cathodic reduction of white phosphorus melt. *Inorganic materials*. **2005**. T.41. №12. P.1463-1467.
- [152] R.A. Lidin, V.A. Molochko, L.L. Andreeva. Chemical properties of inorganic substances: Proc. manual for universities. M: Chemistry. **1996**. P.480.
- [153] S. Rajendran, H. Parveen, K. Begum, R. Chethana. Influence of phosphine on hatching of *Cryptolestes ferrugineus* (Coleoptera: Cucujidae), *Lasioderma serricornis* (Coleoptera: Anobiidae) and *Oryzaephilus surinamensis* (Coleoptera: Silvanidae). *Pest Manag Sci*. **2004**. Vol.60. No.11. P.1114-1118.
- [154] N.S. Nath, I. Bhattacharya, A.G. Tuck, D.I. Schlipalius, P.R. Ebert. Mechanisms of Phosphine Toxicity. *Journal of Toxicology*. **2011**. Vol.2011. P.1-9.
- [155] S. Zuryin, J. Kuang, P. Ebert. Mitochondrial Modulation of Phosphine Toxicity and Resistance in *Caenorhabditis elegans*. *Toxicological Sciences*. **2008**. Vol.102. No.1. P.179-186.
- [156] M.A.G. Pimentel, L.R.D'A. Faroni, R.N.C. Guedes, A.P. Neto, F.M. Garcia. Phosphine resistance, respiration rate and fitness consequences in *Tribolium castaneum* (Herbst) (Coleoptera: Tenebrionidae). *9th International Working Conference on Stored Product Protection. Passo Fundo, RS, Brazil*. **2006**. P.344-351.
- [157] Y.S. Mau, P.J. Collins, G.J. Daghli, M.K. Nayak, H. Pavic, P.R. Ebert. The rph1 Gene Is a Common Contributor to the Evolution of Phosphine Resistance in Independent Field Isolates of *Rhizopertha dominica*. *PLoS ONE*. **2012**. Vol.7. No.2. P.1-13.

- [158] D. Yakhvarov, M. Caporali, L. Gonsalvi, S. Latypov, V. Mirabello, I. Rizvanov, O. Sinyashin, P. Stoppioni, M. Peruzzini. Experimental Evidence of Phosphine Oxide Generation in Solution and Trapping by Ruthenium Complexes. *Angew. Chem. Int. Ed.* **2011**. Vol.50. No.23. P.5370-5373.
- [159] D.G. Yakhvarov, E.V. Gorbachuk, O.G. Sinyashin. Electrode Reactions of Elemental (White) Phosphorus and Phosphane PH₃. *Eur. J. Inorg. Chem.* **2013**. No.27. P.4709-4726.
- [160] J.R. Andreesen. Glycine metabolism in anaerobes. *Antonie van Leeuwenhoek.* **1994**. Vol.66. No.1-3. P.223-237.
- [161] M. Wagner, D. Sonntag, R. Grimm, A. Pich, C. Eckerskorn, B. Söhling, J.R. Andreesen. Substrate-specific selenoprotein B of glycine reductase from *Eubacterium acidaminophilum*. Biochemical and molecular analysis. *Eur. J. Biochem.* **1999**. Vol.260. No.1. P.38-49.
- [162] J.W. Lengeler, G. Drews, H.G. Schlegel. Modern Microbiology. Prokaryotes. V.2. *Moscow: Mir.* **2005**. P.493.
- [163] R.J. Reynolds. The toxicity of phosphine. *Science INTL Report.* **1999**. 28p.
- [164] K. Hemminki. DNA adducts, mutations and cancer. *Carcinogenesis.* **1993**. Vol.14. No.10. P.2007-2012.
- [165] N.K. Kochetkov, E.I. Budovsky, E.D. Sverdlov, N.A. Simukova, M.F. Turchinsky, V.N. Shibaev. Organic chemistry of nucleic acids. *M.: Chemistry.* **1970**. p.720.
- [166] J.A. Swenberg, K. Lu, B.C. Moeller, L. Gao, P.B. Upton, J. Nakamura, T.B. Starr. Endogenous versus Exogenous DNA Adducts: Their Role in Carcinogenesis, Epidemiology, and Risk Assessment. *Toxicological sciences.* **2011**. Vol.120. No.1. P.130-145.
- [167] A. Pähler, J. Parker, W. Dekant. Dose-dependent protein adduct formation in kidney, liver, and blood of rats and human blood after perchloroethene inhalation. *Toxicological Sciences.* **1999**. Vol.48. No.1. P.5-13.
- [168] R.C. Gupta, A. Garg, K. Earley, S.C. Agarwal, G.R. Lambert, S. Nesnow. DNA Adducts of the Antitumor Agent Diaziquone. *Cancer Research.* **1991**. Vol.51. No.19. P.5198-5204.
- [169] J.P. Uittenbogaard, B. Zomer, P. Hoogerhout, B. Metz. Reactions of β -Propiolactone with Nucleobase Analogues, Nucleosides, and Peptides. Implications for the inactivation of viruses. *The journal of biological chemistry.* **2011**. Vol.286. No.42. P.36198-36214.
- [170] L.A. Wessjohann, W. Brandt. Biosynthesis and Metabolism of Cyclopropane Rings in Natural Compounds. *Chem. Rev.* **2003**. Vol.103. No.4. P.1625-1647.
- [171] D.M. Potter, M.S. Baird. Carcinogenic effects of ptaquiloside in bracken fern and related compounds. *British Journal of Cancer.* **2000**. Vol.83. No.7. P.914-920.
- [172] P.J. Brooks, S. Zakhari. Acetaldehyde and the Genome: Beyond Nuclear DNA Adducts and Carcinogenesis. *Environmental and Molecular Mutagenesis.* **2014**. Vol.55. No.2. P.77-91.
- [173] R. Kasiviswanathan, I.G. Minko, R. S. Lloyd, W.C. Copeland. Translesion Synthesis Past Acrolein-derived DNA Adducts by Human Mitochondrial DNA Polymerase γ^* . *Journal of biological chemistry.* **2013**. Vol.288. No.20. P.14247-14255.
- [174] I.G. Minko, I.D. Kozekov, T.M. Harris, C.J. Rizzo, R.S. Lloyd, M.P. Stone. Chemistry and Biology of DNA Containing 1,N²-Deoxyguanosine Adducts of the α,β -Unsaturated Aldehydes Acrolein, Crotonaldehyde, and 4-Hydroxynonenal. *Chem. Res. Toxicol.* **2009**. Vol.22. No.5. P.759-778.
- [175] M. Kawaguchi-Niida, N. Shibata, S. Morikawa, K. Uchida, T. Yamamoto, T. Sawada, M. Kobayashi. Crotonaldehyde accumulates in glial cells of Alzheimer's disease brain. *Acta Neuropathol.* **2006**. Vol.111. P.422-429.
- [176] H. Kasai, N. Iwamoto-Tanaka, S. Fukada. DNA modifications by the mutagen glyoxal: adduction to G and C, deamination of C and GC and GA cross-linking. *Carcinogenesis.* **1998**. Vol.19. No.8. P.1459-1465.
- [177] L.J. Marnett. Lipid peroxidation – DNA damage by malondialdehyde. *Mutation Research.* **1999**. Vol.424. No.1-2. P.83-95.
- [178] R. Olsen, J. Backman, P. Molander, K.D. Klika, L. Kronberg. Identification of Adducts Formed in the Reactions of 2'-Deoxyguanosine and Calf Thymus DNA with Glutaraldehyde. *Eur. J. Org. Chem.* **2007**. Vol.24. P.4011-4018.
- [179] L.S. Von Tungeln, P. Yi, T.J. Bucci, V.M. Samokyszyn, M.W. Chou, F.F. Kadlubar, P.P. Fu. Tumorigenicity of chloral hydrate, trichloroacetic acid, trichloroethanol, malondialdehyde, 4-hydroxy-2-nonenal, crotonaldehyde, and acrolein in the B6C3F₁ neonatal mouse. *Cancer Letters.* **2002**. Vol.185. No.1. P.13-19.
- [180] R.M. LoPachin, A.P. DeCaprio. Protein Adduct Formation as a Molecular Mechanism in Neurotoxicity. *Toxicological Sciences.* **2005**. Vol.86. No.2. P.214-225.
- [181] R.G. Harvey, Q. Dai, C. Ran, K. Lim, I. Blair, T.M. Penning. Syntheses of adducts of active metabolites of carcinogenic polycyclic aromatic hydrocarbons with 2-deoxyribonucleosides. *Polycyclic Aromatic Compounds.* **2005**. Vol.25. No.5. P.371-391.

- [182] D.J. McCarthy, W.R. Waud, R.F. Struck, D.L. Hill. Disposition and Metabolism of Aniline in Fischer 344 Rats and C57BL/6 x C3H F₁ Mice. *Cancer research*. **1985**. Vol.45. No.1. P.174-180.
- [183] M. Murata, S. Kawanishi. Mechanisms of oxidative DNA damage induced by carcinogenic arylamines. *Front Biosci*. **2011**. Vol.16. P.1132-1143.
- [184] V.M. Arlt, M. Stiborova, H.H. Schmeiser. Aristolochic acid as a probable human cancer hazard in herbal remedies: a review. *Mutagenesis*. **2002**. Vol.17. No.4. P.265-277.
- [185] D. Venugopal, M. Zahid, P.C. Mailander, J.L. Meza, E.G. Rogan, E.L. Cavalieri, D. Chakravarti. Reduction of estrogen-induced transformation of mouse mammary epithelial cells by N-acetylcysteine. *Journal of Steroid Biochemistry & Molecular Biology*. **2008**. Vol.109. No.1-2. P.22-30.
- [186] H. Glatt, U. Pabel, W. Meinel, H. Frederiksen, H. Frandsen, E. Muckel. Bioactivation of the heterocyclic aromatic amine 2-amino-3-methyl-9H-pyrido[2,3-b]indole (MeAαC) in recombinant test systems expressing human xenobiotic-metabolizing enzymes. *Carcinogenesis*. **2004**. Vol.25. No.5. P.801-807.
- [187] M.S. Solomon, P.-M. L. Morgenthaler, R.J. Turesky, J.M. Essigmann. Mutational and DNA Binding Specificity of the Carcinogen 2-Amino-3,8-dimethylimidazo[4,5-f]quinoxaline. *The journal of biological chemistry*. **1996**. Vol.271. No.31. P.18368-18374.
- [188] G. Nauwelaers, E.E. Bessette, D. Gu, Y. Tang, J. Rageul, V. Fessard, J.-M. Yuan, M.C. Yu, S. Langouët, R.J. Turesky. DNA Adduct Formation of 4-Aminobiphenyl and Heterocyclic Aromatic Amines in Human Hepatocytes. *Chem. Res. Toxicol*. **2011**. Vol.24. No.6. P.913-925.
- [189] M.C. Byrns, C.C. Vu, J.W. Neidigh, J.-L. Abad, R.A. Jones, L.A. Peterson. Detection of DNA adducts derived from the reactive metabolite of furan, *cis*-2-butene-1,4-dial. *Chem Res Toxicol*. **2006**. Vol.19. No.3. P.414-420.
- [190] J.J. Solomon, A. Segal. DNA Adducts of Propylene Oxide and Acrylonitrile Epoxide: Hydrolytic Deamination of 3-Alkyl-dCyd to 3-Alkyl-dUrd. *Environmental Health Perspectives*. **1989**. Vol.81. P.19-22.
- [191] T.J. Monks, M. Butterworth, S.S. Lau. The fate of benzene-oxide. *Chemico-Biological Interactions*. **2010**. Vol.184. No.1-2. P.201-206.
- [192] M.T. Velasquez, A. Ramezani, D.S. Raj. Urea and protein carbamylation in ESRD: surrogate markers or partners in crime? *Kidney International*. **2015**. Vol.87. No.6. P.1092-1094.
- [193] A. Beyerbach, P.B. Farmer, G. Sabbioni. Biomarkers for Isocyanate Exposure: Synthesis of Isocyanate DNA Adducts. *Chem. Res. Toxicol*. **2006**. Vol.19. No.12. P.1611-1618.
- [194] F.-L. Chung, H.-J.C. Chen, R.G. Nath. Lipid peroxidation as a potential endogenous source for the formation of exocyclic DNA adducts. *Carcinogenesis*. **1996**. Vol.17. No.10. P.2105-2111.
- [195] C. Mesaros, I.A. Blair. Targeted Chiral Analysis of Bioactive Arachidonic Acid Metabolites Using Liquid-Chromatography-Mass Spectrometry. *Metabolites*. **2012**. Vol.2. No.2. P.337-365.
- [196] H. Bartsch, J. Nair. Chronic inflammation and oxidative stress in the genesis and perpetuation of cancer: role of lipid peroxidation, DNA damage, and repair. *Langenbecks Arch Surg*. **2006**. Vol.391. No.5. P.499-510.
- [197] S.A. Marques, A.P.M. Loureiro, O.F. Gomes, C.C.M. Garcia, P. Di Mascio, M.H.G. Medeiros. Induction of 1,N²-etheno-2'-deoxyguanosine in DNA exposed to L-carotene oxidation products. *FEBS Letters*. **2004**. Vol.560. No.1-3. P.125-130.
- [198] S.H. Lee, I.A. Blair. Oxidative DNA Damage and Cardiovascular Disease. *Trends Cardiovasc Med*. **2001**. Vol.9. No.3,4. P.148-155.
- [199] M. Goggin, J.A. Swenberg, V.E. Walker, N. Tretyakova. Molecular Dosimetry of 1,2,3,4-Diepoxybutane-Induced DNA-DNA Cross-Links in B6C3F₁ Mice and F344 Rats Exposed to 1,3-Butadiene by Inhalation. *Cancer Res*. **2009**. Vol.69. No.6. P.2479-2486.
- [200] H.E. Hurst. Toxicology of 1,3-Butadiene, Chloroprene, and Isoprene. *Reviews of Environmental Contamination and Toxicology*. **2007**. Vol.189. P.131-179.
- [201] I. Linhart, P. Mikeš, E. Frantík, J. Mráz. DNA Adducts Formed from p-Benzoquinone, an Electrophilic Metabolite of Benzene, Are Extensively Metabolized in Vivo. *Chem. Res. Toxicol*. **2011**. Vol.24. No.3. P.383-391.
- [202] B. Hang. Formation and Repair of Tobacco Carcinogen-Derived Bulky DNA Adducts. *Journal of Nucleic Acids*. **2010**. Vol.2010. No.709521. 29p.
- [203] M. Murata, R. Thanan, N. Ma, S. Kawanishi. Role of Nitritative and Oxidative DNA Damage in Inflammation-Related Carcinogenesis. *Journal of Biomedicine and Biotechnology*. **2012**. Vol.2012. P.1-11.
- [204] E. Gottschalg, G.B.Scott, P.A.Burns, D.E.G.Shuker. Potassium diazoacetate-induced p53 mutations in vitro in relation to formation of O6-carboxymethyl- and O6-methyl-20-deoxyguanosine DNA adducts: relevance for gastrointestinal cancer. *Carcinogenesis*. **2007**. Vol.28. No.2. P.356-362.

- [205] O.S. Sohn, E.S. Fiala, S.P. Requeijo, J.H. Weisburger, F.J. Gonzalez. Differential Effects of CYP2E1 Status on the Metabolic Activation of the Colon Carcinogens Azoxymethane and Methylazoxymethanol. *Cancer Res.* **2001**. Vol.61. No.23. P.8435-8440.
- [206] A. Trostchansky, H. Rubbo. Lipid nitration and formation of lipid-protein adducts: biological insights. *Amino Acids.* **2007**. Vol.32. No.4. P.517-522.
- [207] K. Naiman, M. Dračinský, P. Hodek, M. Martínková, H.H. Schmeiser, Eva Frei, M. Stiborová. Formation, Persistence, and Identification of DNA Adducts Formed by the Carcinogenic Environmental Pollutant *o*-Anisidine in Rats. *Toxicological Sciences.* **2012**. Vol.127. No.2. P.348-359.
- [208] D. Melton, C.D. Lewis, N.E. Price, K.S. Gates. Covalent Adduct Formation between the Antihypertensive Drug Hydralazine and Abasic Sites in Double- and Single-Stranded DNA. *Chem. Res. Toxicol.* **2014**. Vol.27. No.12. P.2113-2118.
- [209] M.F. Sadiq, W.M. Owais. Mutagenicity of sodium azide and its metabolite azidoalanine in *Drosophila melanogaster*. *Mutation Research.* **2000**. Vol.469. No.2. P.253-257.
- [210] W.M. Owais. The lack of L-azidoalanine interaction with DNA. *In vitro* studies. *Mutat Res.* **1993**. Vol.302. No.3. P.147-151.
- [211] A.A. Wani, J.K. Sullivan, J. Lebowitz. Inununoassays for carbodiimide modified DNA-detection of unpairing transitions in supercoiled ColEI DNA. *Nucleic Acids Research.* **1989**. Vol.17. No.23. P.9957-9977.
- [212] L.J. Marnett. Oxyradicals and DNA damage. *Carcinogenesis.* **2000**. Vol.21. No.3. P.361-370.
- [213] S.B. Nimse, D. Pal. Free radicals, natural antioxidants, and their reaction mechanisms. *RSC Adv.* **2015**. Vol.5. P.27986-28006.
- [214] A. Cajigas, M. Gayer, C. Beam, J. J. Steinberg. Ozonation of DNA Forms Adducts: A 32P-DNA Labeling and Thin-layer Chromatography Technique to Measure DNA Environmental Biomarkers. *Archives of Environmental Health.* **1994**. Vol.49. No.1. P.25-36.
- [215] Y.-C. Yang, J. Yan, D.R. Doerge, P.-C. Chan, P.P. Fu, M.W. Chou. Metabolic Activation of the Tumorigenic Pyrrolizidine Alkaloid, Riddelliine, Leading to DNA Adduct Formation in Vivo. *Chem. Res. Toxicol.* **2001**. Vol.14. No.1. P.101-109.
- [216] B. Prokopczyk, P. Bertinato, D. Hoffmann. Cyanoethylation of DNA *in Vivo* by 3-(Methylnitrosamino)propionitrile, an *Areca*-derived Carcinogen. *Cancer Research.* **1988**. Vol.48. No.23. P.6780-6784.
- [217] W. Tuntiwechapikul, W.M. David, D. Kumar, M. Salazar, S.M. Kerwin. DNA Modification by 4-Aza-3-ene-1,6-diyne: DNA Cleavage, pH-Dependent Cytosine-Specific Interactions, and Cancer Cell Cytotoxicity. *Biochemistry.* **2002**. Vol.41. No.16. P.5283-5290.
- [218] C.L. Sherman, S.E. Pierce, J.S. Brodbelt, B. Tuesuwan, S.M. Kerwin. Identification of the Adduct Between a 4-Aza-3-ene-1,6-diyne and DNA Using Electrospray Ionization Mass Spectrometry. *J Am Soc Mass Spectrom.* **2006**. Vol.17. No.10. P.1342-1352.
- [219] V. Smith, A. Bochkov, R. Capel. Organic synthesis. Science and art. *Moscow: Mir.* **2001**. P.573.
- [220] F.P. Guengerich. Cytochrome P450s and Other Enzymes in Drug Metabolism and Toxicity. *The AAPS Journal.* **2006**. Vol.8. No.1. P.101-111.
- [221] J. Bargonetti, E. Champeil, M. Tomasz. Differential Toxicity of DNA Adducts of Mitomycin C. *Journal of Nucleic Acids.* **2010**. Vol.2010. No.8. P.1-6.
- [222] Elwood A. Mullins, Rongxin Shi & Brandt F. Eichman. Toxicity and repair of DNA adducts produced by the natural product yatakemycin. *Nature Chemical Biology.* **2017**. DOI: 10.1038/nchembio.2439. P.1-9.
- [223] L. Hajovsky, G. Hu, Y. Koen, D. Sarma, W. Cui, D.S. Moore, J.L. Staudinger, R.P. Hanzlik. Metabolism and Toxicity of Thioacetamide and Thioacetamide S-Oxide in Rat Hepatocytes. *Chem. Res. Toxicol.* **2012**. Vol.25. No.9. P.1955-1963.
- [224] B.H. Mathison, M.L. Taylor, M.S. Bogdanffy. Dimethyl sulfate uptake and methylation of DNA in rat respiratory tissues following acute inhalation. *Fundam. appl. Toxicol.* **1995**. Vol.28. No.2. P.255-263.
- [225] R.L. Lundblad. Chemical Modification of Biological Polymers. *CRC Press.* **2011**. 426p.
- [226] G.A. Sega. A review of the genetic effects of ethyl methanesulfonate. *Mutation Research.* **1984**. Vol.134. No.2-3. P.113-142.
- [227] H.M. Bolt, B. Gansewendt. Mechanisms of carcinogenicity of methyl halides. *Crit Rev Toxicol.* **1993**. Vol.23. No.3. P.237-253.
- [228] C. Thomas, Y. Will, S.L. Schoenberg, D. Sanderlin, D.J. Reed. Conjugative metabolism of 1,2-dibromoethane in mitochondria: disruption of oxidative phosphorylation and alkylation of mitochondrial DNA. *Biochemical Pharmacology.* **2001**. Vol.61. No.5. P.595-603.

- [229] G. Samin, D.B. Janssen. Transformation and biodegradation of 1,2,3-trichloropropane (TCP). *Environ Sci Pollut Res.* **2012.** Vol.19. No.8. P.3067-3078.
- [230] W. Dekant. Chemical-induced nephrotoxicity mediated by glutathione S-conjugate formation. *Toxicology Letters.* **2001.** Vol.124. No.1-3. P.21-36.
- [231] H.H. Landin, D. Segerbäck, C. Damberg, S.Osterman-Golkar. Adducts with haemoglobin and with DNA in epichlorohydrin-exposed rats. *Chem Biol Interact.* **1999.** Vol.117. No.1. P.49-64.
- [232] P. Sund, L. Kronberg. Reaction of epichlorohydrin with adenosine, 2'-deoxyadenosine and calf thymus DNA: Identification of adducts. *Bioorganic Chemistry.* **2006.** Vol.34. No.3. P.115-130.
- [233] P.V. Zimakov, O.N. Dyment, N.A. Bogoslovsky, F.I. Weisberg, Yu.M. Stepanov, N.A. Kolchin, R.Sh. Kazarnovskaya, V.A. Sokolova, Yu.A. Kozlova, Yu.Ts. Vol, N.A. Shishakov. Oxyethylene. *M. : Chemistry.* **1967.** P.320.
- [234] W. Kanhai, M. Koob, W. Dekant, & D. Henschler. Metabolism of ¹⁴C-dichloroethyne in rats. *Xenobiotica.* **1991.** Vol.21. No.7. P.905-916.
- [235] S.E. Sparks, G.B. Quistad, J.E. Casida. Chloropicrin: reactions with biological thiols and metabolism in mice. *Chem Res Toxicol.* **1997.** Vol.10. No.9. P.1001-1007.
- [236] A.M. Bracete, M. Sono, J.H. Dawson. Effects of cyanogen bromide modification of the distal histidine on the spectroscopic and ligand binding properties of myoglobin: magnetic circular dichroism spectroscopy as a probe of distal water ligation in ferric high-spin histidine-bound heme proteins. *Biochim Biophys Acta.* **1991.** Vol.1080. No.3. P.264-270.
- [237] B.H. Monien, K. Herrmann, S. Florian, H. Glatt. Metabolic activation of furfuryl alcohol: formation of 2-methylfuranyl DNA adducts in *Salmonella typhimurium* strains expressing human sulfotransferase 1A1 and in FVB/N mice. *Carcinogenesis.* **2011.** Vol.32. No.10. P.1533-1539.
- [238] A. Unnikrishnan, P.R. Gafken, T. Tsukiyama. Dynamic changes in histone acetylation regulate origins of DNA replication. *Nature Structural & Molecular Biology.* **2010.** Vol.17. P.430-439.
- [239] L. Fabrizi, G.W. Taylor, R.J. Edwards, A.R. Boobis. Adducts of the Chloroform Metabolite Phosgene. *Adv Exp Med Biol.* **2001.** Vol.500. P.129-132.
- [240] J.P. Henderson, J. Byun, J.W. Heinecke. Chlorination of nucleobases, RNA and DNA by myeloperoxidase: a pathway for cytotoxicity and mutagenesis by activated phagocytes. *Redox Rep.* **1999.** Vol.4. No.6. P.319-320.
- [241] E.R. Jamieson, S.J. Lippard. Structure, Recognition, and Processing of Cisplatin-DNA Adducts. *Chem. Rev.* **1999.** Vol.99. No.9. P.2467-2498.
- [242] S. Matsuda, S. Matsui, Y. Shimizu, T. Matsuda. Genotoxicity of colloidal fullerene C₆₀. *Environ Sci Technol.* **2011.** Vol.45. No.9. P.4133-4138.
- [243] G.S. Khan, A. Shah, Z.-ur-Rehman, D. Barker. Chemistry of DNA minor groove binding agents. *Journal of Photochemistry and Photobiology B: Biology.* **2012.** Vol.115. P.105-118.
- [244] V.A. Efimov, S.V. Fedyunin. Cross-Linked Nucleic Acids: Isolation, Structure, and Biological Role. *Biochemistry (Moscow).* **2010.** Vol.75. No.13. P.1606-1627. Original Russian Text V. A. Efimov, S. V. Fedyunin. published in *Uspekhi Biologicheskoi Khimii.* **2010.** Vol.50. P.259-302.
- [245] L.A. Johnson, B. Malayappan, N. Tretyakova, C. Campbell, M.L. MacMillan, J.E. Wagner, P.A. Jacobson. Formation of Cyclophosphamide Specific DNA Adducts in Hematological Diseases. *Pediatr Blood Cancer.* **2012.** Vol.58. No.5. P.708-714.
- [246] Z.S. Al Hakkak. Mutagenicity of phosphine gas in *Drosophila melanogaster*. *Journal of Biological Science Research.* **1988.** Vol.19. P.739-745.
- [247] A. Barbosa, A.M. Bonin. Evaluation of phosphine genotoxicity at occupational levels of exposure in New South Wales, Australia. *Occupational and Environmental Medicine.* **1994.** Vol.51. No.10. P.700-705.
- [248] V.F. Garry, J. Griffith, T.J. Danzl, R.L. Nelson, E.B. Whorton, L.A. Krueger, J. Cervenka. Human Genotoxicity: Pesticide Applicators and Phosphine. *Science.* **1989.** Vol.246. No.4927. P.251-255.
- [249] B.B. Pal, S.P. Bhunya. Mutagenicity testing of a rodenticide, zinctox (zinc phosphide) in a mouse in vivo system. *In Vivo.* **1995.** Vol.9. No.1. P.81-83.
- [250] K.A. Muid, R.M. Shahjahan, R. Begum and R.A. Begum. Zinc Phosphide Induced DNA Damage in the Blood Cells of *Gallus* sp. using Comet DNA Assay. *Am. J. Agri. & Biol. Sci.* **2012.** Vol.7. No.1. P.82-87.
- [251] S.A. Buckler, M. Epstein. Reactions of phosphine with ketones: A route to primary phosphine oxides. *Tetrahedron.* **1962.** Vol.18. No.11. P.1211-1219.
- [252] B. Dayde, C. Pierra, G. Gosselin, D. Surleraux, A.T. Ilagouma, C. Laborde, J.-N. Volle, D. Virieux, J.-L. Pirat. Synthesis of Unnatural Phosphonosugar Analogues. *Eur. J. Org. Chem.* **2014.** Vol.2014. No.6. P. 1333- 1337.
- [253] S.A. Beaton, M.P. Huestis, A. Sadeghi-Khomami, N.R. Thomas, D.L. Jakeman. Enzyme-catalyzed synthesis of isosteric phosphono-analogues of sugar Nucleotides. *Chem. Commun.* **2009.** No.2. P.238-240.

- [254] H.J. Banks. The toxicity of phosphine to insects. In: *Proc. 1st Int. Working Conf. on Stored Product Entomology, Savannah, Georgia, USA. 1975*. P.283-296.
- [255] P.A. Hamilton, T. Murrell. Kinetics and Mechanism of the Reactions of PH₃ with O(³P) and N(⁴S) Atoms. *J. Chem. Soc., Faraday Trans 2. 1985*. Vol.81. P.1531-1541.
- [256] J.R. Robinson, E.J. Bond. The toxic action of phosphine: Studies with ³²PH₃; Terminal residues in biological materials. *Journal of Stored Products Research. 1970*. Vol.6. No.2. P.133-146.
- [257] World Health Organization. Phosphine and selected metal phosphides. *Environmental Health Criteria. Geneva. 1988*. 100p.
- [258] H.L. Holland, M. Carey, S. Kumaresan. Fungal biotransformation of organophosphines. *Xenobiotica. 1993*. Vol.23. No.5. P.519-524.
- [259] R. Sidhu. Influence of the reducing agent triphenylphosphine on cyclooxygenase-1 metabolism of arachidonic acid. *Thesis in partial fulfillment of requirements for the degree of Master of Science in Chemistry. 2010*. 49p.
- [260] V. Domkin, A. Chabes. Phosphines are ribonucleotide reductase reductants that act via C-terminal cysteines similar to thioredoxins and glutaredoxins. *Scientific Reports. 2014*. Vol.7. No.4. P.1-7.
- [261] R.A. Wiley, L.A. Sternson, H.A. Sasame, J.R. Gillette. Enzymatic oxidation of diphenylmethylphosphine and 3-dimethylaminopropylidiphenylphosphine by rat liver microsomes. *Biochemical Pharmacology. 1972*. Vol.21. No.24. P.3235-3247.
- [262] D.I. Schlipalius, N. Valmas, A.G. Tuck, R. Jagadeesan, L. Ma, R. Kaur, A. Goldinger, C. Anderson, J. Kuang, S. Zuryn, Y.S. Mau, Q. Cheng, P.J. Collins, M.K. Nayak, H.J. Schirra, M.A. Hilliard, P.R. Ebert. A Core Metabolic Enzyme Mediates Resistance to Phosphine Gas. *Science. 2012*. Vol.338. No.6108. P.807-810.
- [263] B.A. Seibel, P.J. Walsh. Trimethylamine oxide accumulation in marine animals: relationship to acylglycerol storage. *The journal of experimental biology. 2002*. Vol.205. No.3. P.297-306.
- [264] M. Ansaldi, L. Théraulaz, C. Baraquet, G. Panis, V. Méjean. Aerobic TMAO respiration in *Escherichia coli*. *Molecular microbiology. 2007*. Vol.66. No.2. P.484-494.
- [265] K. Hofmann, E. Hammer, M. Köhler, V. Brüser. Oxidation of triphenylarsine to triphenylarsineoxide by *Trichoderma harzianum* and other fungi. *Chemosphere. 2001*. Vol.44. No.4. P.697-700.
- [266] T.R. Miller, R. Belas. Dimethylsulfoniopropionate Metabolism by *Pfiesteria*-Associated *Roseobacter* spp. *Applied and environmental microbiology. 2004*. Vol.70. No.6. P.338-3391.
- [267] K. Kino, T. Murakami-Nitta, M. Oishi, S. Ishiguro, K. Kirimura. Isolation of Dimethyl Sulfone-Degrading Microorganisms and Application to Odorless Degradation of Dimethyl Sulfoxide. *Journal of bioscience and bioengineering. 2004*. Vol.97. No.1. P.82-84.
- [268] D.R. Boyd, N.D. Sharma, A. W.T. King, S.D. Shepherd, C. C.R. Allen, R.A. Holt, H.R. Luckarift, H. Dalton. Stereoselective reductase-catalysed deoxygenation of sulfoxides in aerobic and anaerobic bacteria. *Org. Biomol. Chem. 2004*. Vol.2. No.4. P.554-561.
- [269] T. Kotani, H. Yurimoto, N. Kato, Y. Sakai. Novel Acetone Metabolism in a Propane-Utilizing Bacterium, *Gordonia* sp. Strain TY-5. *Journal of bacteriology. 2007*. Vol.189. No.3. P.886-893.
- [270] D.J. Arp. Butane metabolism by butane-grown '*Pseudomonas butanovora*'. *Microbiology. 1999*. Vol.145. No.5. P.1173-1180.
- [271] J.H. Enemark, C.G. Young. Bioinorganic Chemistry of Pterin-Containing Molybdenum and Tungsten Enzymes. *Advances in Inorganic Chemistry. 1993*. Vol.40. P.1-88.
- [272] F. Lundquist, N. Tygstrup, K. Winkler, K. Mellempgaard, S. Munck-Petersen. Ethanol metabolism and production of free acetate in the human liver. *Journal of Clinical Investigation. 1962*. Vol.41. No.5. P.955- 961.
- [273] Toxicological profile for toluene. *U.S. Department of Health and Human Services, Public Health Service Agency for Toxic Substances and Disease Registry. 2000*. 357p.
- [274] J.K. Ritter. Roles of glucuronidation and UDP-glucuronosyltransferases in xenobiotic bioactivation reactions. *Chemico-Biological Interactions. 2000*. Vol.129. No.1-2. P.171-193.
- [275] T.I. Kazantseva. Synthesis of phosphines and phosphine oxides based on white phosphorus in overbased systems. Thesis for the degree of candidate of chemical sciences in specialty 02.00.08 Chemistry of organoelement compounds. **2004**. P.158.
- [276] Q. Cheng, N. Valmas, P.E.B. Reilly, P.J. Collins, R. Kopittke, P.R. Ebert. *Caenorhabditis elegans* Mutants Resistant to Phosphine Toxicity Show Increased Longevity and Cross-Resistance to the Synergistic Action of Oxygen. *Toxicological sciences. 2003*. Vol.73. No.1. P.60-65.
- [277] C.-H. Hsu, B.-C. Han, M.-Y. Liu, C.-Y. Yeh, J.E. Casida. Phosphine-induced oxydative damage in rats: attenuation by melatonin. *Free radical biology and medicine. 2000*. Vol.28. No.4. P.636-642.

- [278] P.R. Ortiz de Montellano, J.M. Mathews. Autocatalytic alkylation of the cytochrome P-450 prosthetic haem group by 1-aminobenzotriazole. Isolation of an NN-bridged benzyne-protoporphyrin IX adduct. *Biochem. J.* **1981**. Vol.195. No.3. P.761-764.
- [279] M. Volpato, J. Seargent, P.M. Loadman, R.M. Phillips. Formation of DNA interstrand cross-links as a marker of mitomycin C bioreductive activation and chemosensitivity. *Eur J Cancer.* **2005**. Vol.41. No.9. P.1331-1338.
- [280] G.S. Kumar, R. Lipman, J. Cummings, M. Tomasz. Mitomycin C-DNA adducts generated by DT-diaphorase. Revised mechanism of the enzymatic reductive activation of mitomycin C. *Biochemistry.* **1997**. Vol.36. No.46. P.14128-14136.
- [281] S.S. Pan, P.A. Andrews, C.J. Glover, N.R. Bachur. Reductive activation of mitomycin C and mitomycin C metabolites catalysed by NADPH-cytochrome P-450 reductase and xanthine oxidase. *J Biol Chem.* **1984**. Vol.259. No.2. P.959-966.
- [282] G.-One Ahn, B.J. Martin. Combinations of Hypoxia-Targeting Compounds and Radiation Activated Prodrugs with Ionizing Radiation. P.67-91. Multimodal Concepts for Integration of Cytotoxic Drugs. Ed. L.W. Brady, H.-P. Heilmann, M. Molls. *Springer, Heidelberg.* **2006**. 352p.
- [283] A.I. Kourdioukov, V.F. Khayrutdinov, F.M. Gumerov, A.R. Gabitova, V.G. Uryadov, A.F. Mingaliev, E.N. Ofitserov. Triplet oxygen-water associates as the main agents of acidifying autocatalytic redox-processes. Quantum-chemical description of primary elementary acts of combustion. *Butlerov Communications.* **2017**. Vol.52. No.10. P.17-27. ROI: jbc-02/17-52-10-17
- [284] C. Brunel, A. Bondon, G. Simonneaux. Trimethylphosphine binding to horse-heart and sperm-whale myoglobins. Kinetics, proton magnetic resonance assignment and nuclear Overhauser effect investigation of the heme pocket. *Eur J Biochem.* **1993**. Vol.214. No.2. P.405-414.
- [285] K. Kirk, P.W. Kuchel. Characterization of transmembrane chemical shift differences in the ³¹P NMR spectra of various phosphoryl compounds added to erythrocyte suspensions. *Biochemistry.* **1988**. Vol.27. No.24. P.8795-8802.
- [286] A. Bondon, G. Simonneaux. ³¹P-NMR investigation of trimethylphosphine binding to [alpha Fe(II), beta Mn(II)] hybrid hemoglobin. A model for partially liganded species. *Biophys Chem.* **1990**. Vol.37. No.1-3. P.407-411.
- [287] I. Bertini, G. Gray, E. Stifel, J. Valentine. Biological inorganic chemistry: structure and reactivity. *BINOM. Trans. from English.* **2013**. Vol.1. P.456.
- [288] M. Johnson, J. Thuman, R.G. Letterman, C.J. Stromberg, C.E. Webster, E.J. Heilweil. Time-Resolved Infrared Studies of a Trimethylphosphine Model Derivative of [FeFe]-Hydrogenase. *J. Phys. Chem. B.* **2013**. Vol.117. No.39. P.15792-15803.
- [289] L. Vaska, J.W. DiLuzio. Carbonyl and hydrido-carbonyl complexes of iridium by reaction with alcohols. Hydrido complexes by reaction with acid. *J. Am. Chem. Soc.* **1961**. Vol.83. No.12. P.2784-2785.
- [290] M.P. Rigobello, G. Scutari, R. Boscolo, A. Bindoli. Induction of mitochondrial permeability transition by auranofin, a Gold(I)-phosphine derivative. *British Journal of Pharmacology.* **2002**. Vol.136. No.8. P.1162-1168.
- [291] M. Hughes. Inorganic chemistry of biological processes. *Moscow: Mir.* **1983**. P.4144.
- [292] R.J. Clark, M.A. Busch. Stereochemical studies of metal carbonyl-phosphorus trifluoride complexes. *Acc. Chem. Res.* **1973**. Vol.6. No.7. P.246-252.
- [293] X. Zhu, N.A. Larsen, A. Basran, N.C. Bruce, I.A. Wilson. Observation of an Arsenic Adduct in an Acetyl Esterase Crystal Structure. *The Journal Of Biological Chemistry.* **2003**. Vol.278. No.3. P.2008-2014.
- [294] Yu.S. Kagan. Toxicology of organophosphorus pesticides. *M. : Medicine.* **1977**. P.296
- [295] K. Los. Synthetic poisons. *M. : Foreign Literature Publishing House.* **1963**. P.258.
- [296] G.I. Oxygenendler. Poisons and antidotes. *L. : Science. Leningrad branch.* **1982**. P.191
- [297] S.G. Galaktionov. Biologically active. *M. : Young Guard.* **1988**. P.272.
- [298] Z. Franke, P. Franz, V. Warnke. Chemistry of chemical agents. Translation from German. *M. : Chemistry.* **1973**. V.1. P.440
- [299] Z. Franke, P. Franz, V. Warnke. Chemistry of chemical agents. Trans. with him., Ed. acad. I.L. Knunyants and Dr. Chem. Sciences.R. N. Sterling. *M. : Chemistry.* **1973**. V.2. P.404.
- [300] L.P. Kuznetsova, L.I. Kugusheva, E.B. Nikol'skaia. [Catalytic properties of cholinesterases immobilized in a gelatin membrane]. *Ukr Biokhim Zh.* **1990**. Vol.62. No.6. P.42-48.
- [301] S.A. Kardos, L.G. Sultatos. Interactions of the Organophosphates Paraoxon and Methyl Paraoxon with Mouse Brain Acetylcholinesterase. *Toxicological Sciences.* **2000**. Vol.58. No.1. P.118-126.
- [302] J. Blasiak, P. Jalszynski, A. Trzeciak, K. Szyfter. In vitro studies on the genotoxicity of the organophosphorus insecticide malathion and its two analogues. *Mutat Res.* **1999**. Vol.445. No.2. P.275-283.

- [303] M. Quitschau, T. Schuhmann, J. Piel, P. von Zezschwitz, S. Grond. The New Metabolite (S)-Cinnamoylphosphoramidate from *Streptomyces* sp. and Its Total Synthesis. *Eur. J. Org. Chem.* **2008**. Vol.2008. No.30. P.5117-5124.
- [304] G. Petroianu. Natural phosphor ester cholinesterase inhibitors. *Mil. Med. Sci. Lett. (Voj. Zdrav. Listy)*. **2012**. Vol.81. No.2. P.82-83.
- [305] R.K. Malla, S. Bandyopadhyay, C.D. Spilling, S. Dutta, C.M. Dupureur. The First Total Synthesis of (±)-Cyclophostin and (±)-Cyclipostin P: Inhibitors of the Serine Hydrolases Acetyl Cholinesterase and Hormone Sensitive Lipase. *Org. Lett.* **2011**. Vol.13. No.12. P.3094-3097.
- [306] K.L. Roland, J. W. Little. Reaction of LexA Repressor with Diisopropyl Fluorophosphate. A test of the serine protease model. *The journal of biological chemistry*. **1990**. Vol.265. No.22. P.12828-12835.
- [307] V.C. Akimenko. Polyhydroxyalkanoates of microorganisms: searching for natural and constructing recombinant producer strains, biosynthesis, use. *G.K. Scriabin Institute of Biochemistry and Physiology of Microorganisms of Russian Academy of Sciences. M.: TiRu*. **2015**. P.284.
- [308] R. Jagadeesan, A. Fotheringham P.R Ebert, D.I. Schlipalius. Rapid genome wide mapping of phosphine resistance loci by a simple regional averaging analysis in the red flour beetle, *Tribolium castaneum*. *BMC Genomics*. **2013**. Vol.14. No.650. P.1-12.
- [309] M. Fu, X. Song, Z. Yu, Y. Liu. Responses of Phosphate Transporter Gene and Alkaline Phosphatase in *Thalassiosira pseudonana* to Phosphine. *PLoS ONE*. **2013**. Vol.8. No.3. P.1-9.
- [310] R.D. Woodyer, Z. Shao, P.M. Thomas, N.L. Kelleher, J.A.V. Blodgett, W.W. Metcalf, W.A. van der Donk, H. Zhao. Heterologous Production of Fosfomycin and Identification of the Minimal Biosynthetic Gene Cluster. *Chemistry & Biology*. **2006**. Vol.13. No.11. P.1171-1182.
- [311] H. Seto, T. Kuzuyama. Bioactive natural products with carbon-phosphorus bonds and their biosynthesis. *Nat. Prod. Rep.* **1999**. Vol.16. No.5. P.589-596.
- [312] H.W. van Veen. Phosphate transport in prokaryotes: molecules, mediators and mechanisms. *Antonie van Leeuwenhoek*. **1997**. Vol.72. No.4. P.299-315.
- [313] C. Reichmuth. Uptake of phosphine by stored-product pest insects during fumigation. *Proc. 6th Int. Working Conf. on Stored-Product Protection. Canberra, Australia*. **1994**. P.157-162.
- [314] C. Waterford. Distribution of radio-labelled phosphorus in susceptible and resistant *Tribolium castaneum* after fumigation. *Proc. Int. Conf. Controlled Atmosphere and Fumigation in Stored Products, Fresno, CA*. **2000**. P.71-77.
- [315] R.I. Krieger. Handbook of Pesticide Toxicology. *Academic Press*. **2001**. P.1908. T.W. Clarkson. Chapter 61 – Inorganic and Organometal Pesticides. P.1357-1428.
- [316] S.N. Golikov. Emergency care for acute poisoning. *Medicine*. **1978**. P.312.
- [317] Toxicological profile for white phosphorus. *U.S. Department of health and human services. USA*. **1997**. P.248.
- [318] V.A. Alekseenko, S.A. Buzmakov, M.S. Panin. Geochemistry of the environment. *Publishing house of the Perm State National Research University*. **2013**. P.359
- [319] A.F. Verbovoy. The state of bone tissue and calcium-phosphorus metabolism in phosphorus production workers. *Kazan Medical Journal*. **2002**. Vol.83. №5. P.147-150.
- [320] A.F. Verbovoy. Indicators of phosphorus-calcium metabolism and bone tissue density in phosphorus working in production. *Osteoporosis and osteopathy*. **1999**. Issue 4. P.11-13.
- [321] Yu. Medvedev, E. Basin. Phosphoric necrosis of the jaws. *Doctor*. **2012**. №1. P.21-25.
- [322] J. P. Whalen, N. O'donohue, L. Krook, E. A. Nunez. Pathogenesis of abnormal remodeling of bones: effects of yellow phosphorus in the growing rat. *anat. Rec.* **1973**. Vol.177. No.1. P.15-22.
- [323] D.W. Sparling, D. Day, P. Klein. Acute Toxicity and Sublethal Effects of White Phosphorus in Mute Swans, *Cygnus olor*. *Environmental Contamination and Toxicology*. **1999**. Vol.36. P.316- 322.
- [324] O.U. Fernandez, L.L. Canizares. Acute hepatotoxicity from ingestion of yellow phosphorus-containing fireworks. *JClinGastroenterol*. **1995**. Vol.21. No.2. P.139-42.
- [325] A. Cheng, A. Mavrokokki, G. Carter, B. Stein, N.L. Fazzalari, D.F. Wilson, A.N. Goss. The dental implications of bisphosphonates and bone disease. *Australian Dental Journal Medications Supplement*. **2005**. Vol.50. No.2. P. 4-13.
- [326] J.J. Carey, L. Palomo. Bisphosphonates and osteonecrosis of the jaw: Innocent association or significant risk? *Cleveland clinic journal of medicine*. **2008**. Vol.75. No.12. P.871-879.
- [327] G. Ficarra, F. Beninati. Bisphosphonate – related osteonecrosis of the jaws: the point of view of the oral pathologist. *Clinical Cases in Mineral and Bone Metabolism*. **2007**. Vol.4. No.1. P.53-57.
- [328] R.A. Zingaro, R.E. McGlothlin. Some Phosphines, Phosphine Sulfides, and Phosphine Selenides. *J. Chem. Eng. Data*. **1963**. Vol.8. No.2. P.226-229.

- Review** _____ A.Z. Mindubaev, Akosah Yaw Abayie, and D.G. Yakhvarov
- [329] X. Ye, N. Hayashi, S. Tsuboi. A Novel Synthesis of Symmetric Trienes. *J. Fac. Environ Sci. and Tech., Okayama Univ.* **2000**. Vol.5. No.1. P.153-158.
- [330] García-Álvarez, S.E. García-Garrido, V. Cadierno. Iminophosphorane-phosphines: Versatile ligands or homogeneous catalysis. *Journal of Organometallic Chemistry.* **2014**. Vol.751. P.792-808.
- [331] L.V. Avdeeva, T.L. Aleynikova, L.E. Andrianova, N.N. Belushkina, N.P. Volkova, S.A. Vorobyeva, V.A. Golenchenko, A.E. Gubareva, O.V. Korlyakova, N.V. Likhacheva, N.A. Pavlova, G.V. Rubtsova, S.A. Silaeva, S.N. Siluyanova, T.A. Titova. *Biochemistry: a textbook.* Edited by E.S. Severin. 5th ed., Rev. *Moscow: GEOTAR-Media.* **2013**. P.76.